1	The Brighton Collaboration Standardized Template for Collection of Key Information for
2	Risk/Benefit Assessment of Nucleic Acid (RNA and DNA) Vaccines
3	Denny Kima
4	James S. Robertsonb
5	Jean-Louis Exclerc
6	Richard C Conditd
7	Patricia E. Faste
8	Marc Gurwithf
9	George Pavlakisg
10	Thomas P. Monathh
11	Jonathan Smithi
12	David Woodb
13	Emily Smithf
14	Robert T Chenf
15	Sonali Kochharj
16	For the Brighton Collaboration Viral Vector Vaccines Safety Working Group (V3SWG)2
17	
18	a. Janssen Pharmaceuticals, Titusville, NJ, USA
19	b. Independent Adviser, United Kingdom
20	c. International Vaccine Institute, Seoul, Republic of Korea
21	d. Department of Molecular Genetics and Microbiology, University of Florida, Gainesville, FL
22	32610, USA.

- 23 e. International AIDS Vaccine Initiative, New York, NY 10004 USA; Stanford School of
- 24 Medicine, Paolo Alto, CA 94305, USA
- 25 f. Brighton Collaboration, a program of the Task Force for Global Health, Decatur, GA, USA
- 26 g. National Cancer Institute, National Institutes of Health, Frederick MD 21702 USA
- 27 h. Crozet BioPharma LLC, Devens, MA 01434 USA
- i. VLP Therapeutics, Gaithersburg, MD 20878 USA
- 29 j. Global Healthcare Consulting, New Delhi, India; University of Washington, Seattle, WA 98195,
- 30 USA

31

- 32 1. Corresponding author: email address: brightoncollaborationv3swg@gmail.com
- 33 2. See Acknowledgement for other V3SWG members

34

- 35 Disclaimer: The findings, opinions, conclusions, and assertions contained in this consensus
- document are those of the individual members of the Working Group. They do not necessarily
- 37 represent the official positions of any participant's organization (e.g., government, university, or
- 38 corporations) and should not be construed to represent any Agency determination or policy.

39

- 40 Acknowledgment: We thank the following colleagues for their helpful advice: (1) Margaret Liu,
- 41 Brighton Collaboration members; and (2) other members of the V3SWG during the preparation of
- 42 this document: Karin Bok, Najwa Khuri-Bulos, Bettina Klug, and Merita Kucuku.

43

44

Abstract:

- Nucleic acid (DNA and RNA) vaccines are among the most advanced vaccines for COVID-19 under development. The Brighton Collaboration Viral Vector Vaccines Safety Working Group (V3SWG) has prepared a standardized template to describe the key considerations for the risk-benefit assessment of nucleic acid vaccines. This will facilitate key stakeholders to anticipate potential safety issues and interpret or assess safety data. This would also help improve communication and public acceptance of licensed nucleic acid vaccines.
- **Key Words:**
- Brighton Collaboration, COVID-19, CEPI, vaccines, risk-benefit, nucleic acid
- **Introduction:**

The Brighton Collaboration (www.brightoncollaboration.org) was launched in 2000 to improve the science of vaccine safety. The Brighton Collaboration formed the Viral Vector Vaccines Safety Working Group (V3SWG) in October 2008 to improve the ability of key stakeholders to anticipate potential safety issues and meaningfully assess or interpret safety data, thereby facilitating greater public acceptance when viral vector vaccines are licensed. The V3SWG has since published completed standardized templates describing the key considerations for a risk-benefit assessment of several new viral vectors or their vaccines. The information on the template will hopefully facilitate communication of otherwise complex and highly technical data among key stakeholders (some of whom may lack subspecialized training in biotechnology) and increase the transparency, comparability, and comprehension of essential information. The template has been used for the standardized risk-assessment of several new viral vector vaccines,3-5 including some targeting Ebola. The WHO Global Advisory Committee on Vaccine Safety (GACVS)

endorsed the use of the template for other new candidate Ebola vaccines "as it is a structured approach to vaccine safety".6

In 2020, the development of vaccines for COVID-19 is appropriately occurring with unprecedented speed.7 The pace and volume of development make a deliberate and systematic approach that is accessible and understandable to a diversity of stakeholders all the more important. Several DNA and RNA vaccine candidates are among the most advanced COVID-19 vaccines in development. The Brighton Collaboration V3SWG has therefore developed a specific template for nucleic acid vaccines that the Coalition for Epidemic Preparedness Innovations (CEPI) and other key stakeholders will use to evaluate and communicate the benefit-risk of vaccines using these nucleic acid platforms. See Supplementary Material for definitions and additional guidance for completing this template.

DNA vaccines have been under development since the early 1990's. They comprise a bacterial plasmid DNA expressing an immunogen of interest under the control of a eukaryotic promoter. This results in the *de novo* synthesis of the immunogen in the vaccine recipient and the stimulation of both B and T cell immune responses. DNA vaccination was a highly promising approach to vaccination with relatively straightforward construction of the vaccine and ease of large-scale manufacture. Some are licensed for veterinary use and some have undergone clinical trials in humans, but to date none are licensed in humans. Due to the very low immune response in humans with simple naked plasmid DNA, research has focused on methods to enhance the response, including optimizing codon usage, optimizing the formulation for improved uptake of the DNA, optimizing the route or method of administration, or the co-administration of DNA encoding

immune stimulatory molecules. The use of DNA to prime an individual followed by a heterologous vaccination with the same antigen in an alternate format, e.g. a viral vector, is producing promising results. Due to the uniqueness of DNA as a vaccine and the approaches being used to improve their immunogenic effect, vaccination with DNA presents a unique set of safety issues.8 The 2019 proposed revision of the WHO guidelines on DNA vaccines lists the approaches being employed to enhance the immunogenicity of a DNA vaccine.9

RNA vaccines are a more novel approach. An RNA vaccine is typically a messenger RNA molecule that encodes the immunogen of interest; some RNA vaccines employ self-amplifying RNA that directs its own replication within the host cell thus expressing more of the immunogen. Self-amplifying RNA vaccines typically link the antigen-encoding RNA to an RNA replication cassette derived from an RNA virus. None have been licensed for use in either humans or animals, but several have shown promise in animal models and one is currently undergoing Phase I clinical trials. 10 In contrast to a DNA vaccine, an RNA vaccine is translated directly within the cytoplasm of the cell without the need to be transported into the nucleus for transcription; thus there is no concern regarding insertional mutagenesis. Similar to a DNA vaccine though, the *de novo* intracellular synthesis of the immunogen of an RNA vaccine stimulates both B and T cell responses. Due to the greater lability of RNA compared with DNA, more care has to be given to their formulation. More data is required on RNA vaccines safety profile.11,12

RNA and DNA vaccines have, in theory, a distinct advantage of rapid development and deployment, especially in the context of an emerging pandemic, because the only requirement for

114	construction of any particular vaccine is the nucleic acid sequence of the immunodominant
115	antigen(s) of the target pathogen.
116	
117	
118	
119	
120	
121	
122	
123	
124	
125	
126	
127	
128	
129	
130	
131	
132	
133	
134	
135	
136	

Supplementary Material

Specific Instructions for Completing the V3SWG Template:

 Please read these instructions before you complete the nine sections. Send questions to:brightoncollaborationv3swg@gmail.com

• The first section entitled "Authorship" should include your name and the latest date completing the form. If you are working with someone else to complete this form, their name should be provided as well. If you are updating the form, please provide the updated date. These co-authors will be included in the final published template in Vaccine once reviewed and approved by the V3SWG and in subsequent Wiki updates on the V3SWG website.

• Sections 2-7 collect information regarding the basic vaccine information (Section 2), the target pathogen and population (Sections 3), characteristics of transgene and expression, (Section 4), delivery and administration (Section 5), toxicology and nonclinical (Section 6) and human efficacy and other important information (Section 7). Depending on the vaccine, some sections may be redundant or not applicable, for example if the section is for a DNA vaccine but the template is being completed for a RNA vaccine. In cases of redundancies, an answer may simply refer to the answer in a previous section.

• Answer questions by responding in the column entitled 'Information.' If you have any comments or concerns regarding the question or your answer to the question, note these in the 'Comments/Concerns' column. Finally, please provide references in the 'Reference' column. More than one reference can be used per question. You can simply write the first author's last name, first name initials, and year of publication (e.g. Lewis MH, 2003) in the "Reference" column here, but please provide the full citation for the reference at the end of the form. Unpublished data are acceptable, though we do wish for you to include the source and contact information.

• Sections 8 and 9 have column titles that differ from preceding sections intended to provide a summary assessment of adverse effects and toxicity of the vaccine. Please summarize adverse effects and toxicities as requested and rate the risk in the following fashion: none, minimal, low, moderate, high, or unknown. If there is insufficient data for use of the platform in humans to accurately make these assessments, please state so in response to the questions.

 When completing information on adverse effects in Section 8, please provide as many details as possible based on the Brighton Collaboration Guidelines for collection, analysis and presentation of vaccine safety data in pre- and post-licensure clinical studies.¹³

• If a literature search was conducted to complete any of the Sections (strongly encouraged), please add the following information in the Reference(s) column: 1) time

181	period covered (e.g., month/year to month/year); 2) Medical Subject Headings (MeSH)
182	terms used; 3) the number of references found; and 4) the actual references with relevant
183	information used. For prior published templates, please search PubMed for "Brighton
184	Collaboration V3SWG".
185	
186	
187	
188	
189	
190	
191	
192	
193	
194	
195	
196	
197	
198	
199	
200	
201	
202	
203	

204 References

205

- 206 1. Bonhoeffer J, Kohl K, Chen R, Duclos P, Heijbel H, Heininger U et al. The Brighton
- 207 Collaboration: addressing the need for standardized case definitions of adverse events following
- 208 immunization (AEFI). Vaccine. 2002;21(3-4):298-302. PubMed PMID:
- 209 12450705. doi.org/10.1016/S0264-410X(02)00449-8

210

- 2. Chen RT, Carbery B, Mac L, Berns KI, Chapman L, Condit RC et al. The Brighton
- 212 Collaboration Viral Vector Vaccines Safety Working Group (V3SWG). Vaccine. 2015; 33(1):73-
- 213 5. doi:10.1016/j.vaccine.2014.09.035

214

- 3. Monath TP, Seligman SJ, Robertson JS, Guy B, Hayes EB, Condit RC et al. Live virus vaccines
- based on a yellow fever vaccine backbone: standardized template with key considerations for a
- 217 risk/benefit assessment. Vaccine. 2015;33(1):62–72. doi:10.1016/j.vaccine.2014.10.004

218

- 4. Clarke DK, Hendry RM, Singh V, Rose JK, Seligman SJ, Klug B et al. Live virus vaccines
- 220 based on a vesicular stomatitis virus (VSV) backbone: Standardized template with key
- 221 considerations for a risk/benefit assessment. Vaccine. 2016;34(51):6597–6609.
- doi:10.1016/j.vaccine.2016.06.071

- 224 5. Monath TP, Fast PE, Modjarrad K, Clarke DK, Martin BK, Fusco J et al. rVSVΔG-ZEBOV-
- GP (also designated V920) recombinant vesicular stomatitis virus pseudotyped with Ebola Zaire

226 Glycoprotein: Standardized template with key considerations for a risk/benefit assessment. 227 Vaccine X. 2019; 1:100009. doi: 10.1016/j.jvacx.2019.100009 228 229 6. World Health Organization. Global Advisory Committee on Vaccine Safety, 230 4–5 December 2019: Ad26.ZEBOV/MVA-BN-Filo vaccine. Wkly Epidem Rec 2020; 95:28–30 231 232 7. Thanh Le T, Andreadakis Z, Kumar A, Gómez Román R, Tollefsen S, Saville M et al. The 233 COVID-19 vaccine development landscape [published online ahead of print, 2020 Apr 9]. Nat Rev 234 Drug Discov. 2020;10.1038/d41573-020-00073-5. doi:10.1038/d41573-020-00073-5 235 236 8. Liu MA. A Comparison of Plasmid DNA and mRNA as Vaccine Technologies. Vaccines 237 (Basel). 2019;7(2). pii: E37. doi: 10.3390/vaccines7020037 238 239 9. Guidelines for assuring the quality, safety, and efficacy of DNA vaccines. Proposed revision of 240 Annex 1 of WHO Technical Report Series, No. 941. Draft 26 July 2019. WHO. Accessed on April 241 8, 2020 at http://www9.who.int/biologicals/WHO_DNA_vaccine_HK_26_July_2019.pdf 242 243 10. Safety and Immunogenicity Study of 2019-nCoV Vaccine (mRNA-1273) for Prophylaxis 244 SARS-CoV-2 Infection (COVID-19). Clinicaltrials.gov. Link: 245 https://clinicaltrials.gov/ct2/show/NCT04283461 246

247 11. Pardi N, Hogan MJ, Porter FW, Weissman D. mRNA vaccines - a new era in vaccinology. Nat

248 Rev Drug Discov. 2018; 17(4):261-279. doi: 10.1038/nrd.2017.243

249								
250	12. Iavar	rone C, O'Hag	an DT, Yu	D, Delah	aye NF, Ulmer	JB. Mecha	nism of action of m	ıRNA-
251	based	vaccines.	Expert	Rev	Vaccines.	2017;	16(9):871-881.	doi:
252	10.1080/	14760584.201	7.1355245					
253								
254	13. Bonh	noeffer J, Bent	si-Enchill A	, Chen R'	Γ, Fisher MC, 0	Gold MS, H	artman K et al. Guio	delines
255	for collec	ction, analysis	and present	ation of v	vaccine safety of	data in pre-	and post-licensure c	linical
256	studies.	Vaccine. 2009;	;27(16):2282	2-8. doi.o	rg/10.1016/j.va	ccine.2008.	11.036.	
257								
258 259 260 261	Declarat	tion of Intere	st: No auth	ors have	known comp	peting finan	cial interests or pe	ersonal
262	relations	hips that could	l have appea	red to inf	luence the worl	k reported in	n this paper.	
263								
264								
265								
266								
267								
268								
269								
270								
271								

Version date: May 1, 2020 Table 1.

272

Brighton Collaboration

Concatenated Version of Standardized Template for Collection of Key Information for Risk Assessment of Nucleic Acid (RNA and DNA) Vaccines.

See for regular https://brightoncollaboration.us/v3swg/ version.

1. Authorship	2. Basic Vaccine information	3. Target Pathogen and Population	4. Characteristics of Vaccine Transgene and Expression	5. Delivery and Administration	6. Toxicology and Nonclinical	7. Human Efficacy and Other Important Information	8. Adverse Event (AE) Assessment of the Vaccine Platform (*see Instructions):	9. Overall Risk Assessment
1.1. Author(s)	2.1 Vaccine name	3.1 What is the target pathogen?	4.1 Nature of the nucleic acid platform (DNA - synthetic, bacterial, plasmid, linear, >1 type/molecule, other; RNA-messenger, self-replicating, other)	5.1 How might the delivery vehicle impact the safety profile of the vaccine?	6.1 How long does the RNA or DNA persist in vivo (may specify in tissue/serum, proximal/dist al to site of administratio n)?	7.1 What is the evidence that the vaccine would generate a protective immune response in humans (e.g., natural history, passive immunizati on, animal challenge studies)?	8.1. Approximately how many humans have received this vaccine to date? If variants of the vaccine platform, please list separately	9.1 Please summarize key safety issues of concern identified to date, if any:

Version date: May 1, 2020

1	Ī	I	ī	ı	ī	ı	Version	1 date: May 1, 2020
1.2. Date completed/upda ted	2.2 Nucleic Acid Type: DNA, RNA, self- amplifying RNA	3.2 What are the disease manifestations caused by the target pathogen in humans, for the following categories:	4.2 Gene(s) incorporated into the vaccine (antigen, T-cell epitopes, antibiotic resistance factors, cytokines, other)	5.2 How might the vaccine delivery method impact the safety profile?* (e.g., electroporation (please specify name of device), intradermal needle injection)	6.2 What is the risk of integration of sequences from the platform nucleic acid into the host genome?	7.2 Describe other key information that may impact Benefit-Risk	8.2. Method(s) used for safety monitoring:	•how should they be addressed going forward
	2.3 Adjuvant (if applicable)	●In healthy people	4.3 Factors enhancing/controll ing gene expression	5.3 How might any co-administered components (e.g., adjuvants, cytokines, immunomodulat ory molecules) impact the safety profile?	6.3 What is the possible risk of autoimmunity or a harmful immune response?		•Spontaneous reports/passive surveillance	9.2 What is the potential for causing serious unwanted effects and toxicities in:
	2.4 Final Vaccine Formulation: Identify components of formulation that may impact delivery into cells, stability, and safety (e.g., complexing with polymers, encapsulation within microparticle s, liposomes)	●In immunocompromi sed people	4.4 Non-expressed features impacting vaccine efficacy (CpG sequences, other)	5.4 If applicable, describe the heterologous prime-boost regimen that this vaccine is a part of and the possible impact on safety	6.4 Do animal models for toxicity exist? Summarize results		●Diary	• healthy humans?

Version	date:	May	1, 2020

2.5 Route and method of Delivery (e.g., IM injection, gene gun, electroporati on)	•In neonates, infants, children	4.5 Other sequence features that may impact safety (e.g., sequences in DNA that might facilitate insertion or recombination)	5.5 Describe how components of formulation may impact delivery into cells, stability**, and safety (e.g., complexing with polymers, encapsulation within microparticles, nanoparticles and liposomes)	6.5 Do animal models for immunogenic ity and efficacy exist? Summarize results	• Other active surveillance	•immunocompromi sed humans?
	•During pregnancy and in the fetus	4.6 Is the transgene likely to induce immunity to all strains/genotypes of the target pathogen?		6.6 What is the evidence of disease enhancement (if any) in vitro or in animal models?	8.3. What criteria were used for grading the AEs?	•human neonates, infants, children?
	●In elderly	4.7 What is known about the immune response to the vaccine (binding, functional, and neutralizing antibody, B-cell, T-cell memory, etc.)?		6.7 What is known about biodistributio n of the platform nucleic acid in animal models?	•2007 US FDA Guidance for Industry Toxicity Grading Scale for Healthy Adult and Adolescent Volunteers Enrolled in Preventive Vaccine Clinical Trials	•pregnancy and in the unborn in humans?
	•In any other special populations			6.8 Does the vaccine have any impact on innate immunity? If so, what are the implications	• If no criteria were used for grading, or if other metrics were employed, please describe:	•in any other special populations

			 Version	n date: May 1, 2020
		for Benefit- Risk?		
a a e e c c c c c c c c c c c c c c c c	3.3 Briefly, what are the key epidemiologic characteristics of the disease caused by the target pathogen (e.g., incubation period, communicable period, route/s of transmission, case fatality rate, transmissibility characteristics such as basic reproductive ratio (R ₀))?	6.9 What is the evidence that the vaccine has generated a beneficial immune response in:	8.4. List and provide frequency of any related or possibly related serious* AEs observed (*see Instructions)	
o and b p p p p p w w fee	3.4 What sections of the population are most affected by the target pathogen (e.g., pediatric, pregnant, lactating women (breastfeeding), adult, elderly)?	•Small animal models?	8.5. List and provide frequency of any serious, unexpected AE	

Version date: May 1, 2020 **8.6.** List and provide frequency **3.5** What is known of any serious, about the unexpected Nonhuman correlates of statistically primates protective significantly immunity to the (NHP)? increased AE or target pathogen or lab abnormality in to the disease? vaccinee vs. control group **3.6** Please describe any other key information •Describe the about the target control group: pathogen or population that may inform benefit risk **8.7.** List and provide frequency of Adverse Events of Special Interest **8.8.** What is the evidence of disease enhancement (if any) in humans? **8.9.** Did a Data Safety Monitoring Board (DSMB) or its equivalent oversee the study? •Did it identify any safety issue of concern? •If so describe

^{*} Also consider the safety impact of multi-dose delivery methods, the use of multi dose vaccine vials, and any special considerations for disposal.

^{**} Stability is considered here in the context of any relevant intrinsic characteristic of the vaccine deemed relevant for safety purposes. For example, among the risks that WHO, FDA, and EMA list for the use of DNA vaccines is the hazard of integration into recipient's chromosomal DNA with the resulting risk of insertional mutagenesis or spreading of antibiotics resistance genes. The probability of chromosomal integration increases if the introduced pDNA has been linearized, and this is the reason that

Version date: May 1, 2020

regulatory authorities require the plasmid preparation intended for vaccination or gene therapy to contain a high percentage of supercoiled material (usually >80%). The percentage of supercoiled material is also used as a criterion of DNA vaccine stability at different storage temperatures.